

# Comprehensive evaluation the stability of internal control genes for qRT-PCR normalization in cuttlefish *Sepiella japonica* using geNorm, NormFinder, and BestKeeper

Tel: 18025353742 E-mail:: 1435483833@qq.com 谷敏,崔闻博,刘燕琳,周旭,李双\*,迟长凤\* 浙江海洋大学海洋科学与技术学院,海洋生物种质资源发掘利用国家地方 联合工程研究中心,浙江舟山,316022

Abstract: Cuttlefish, Sepiella japonica, is an economically important cephalopoda species in East China Sea. Recently, a growing number of publications on biological research of this species have been made, and many of them were dealing with gene expression analysis. Quantitative real-time PCR (qRT-PCR) is a widely used method for analyzing gene expression, with its accuracy heavily reliant on selecting appropriate reference genes for normalization. So far, no studies on internal control gene screening have been reported in cuttlefish. Therefore, to obtain reliable results from qPCR analysis, six commonly used candidate reference genes were assessed in various tissues in both male and female cuttlefish, namely  $EF-1\gamma$ ,  $EF-1\alpha$ , E

### 1) Expression profiling of candidate reference genes

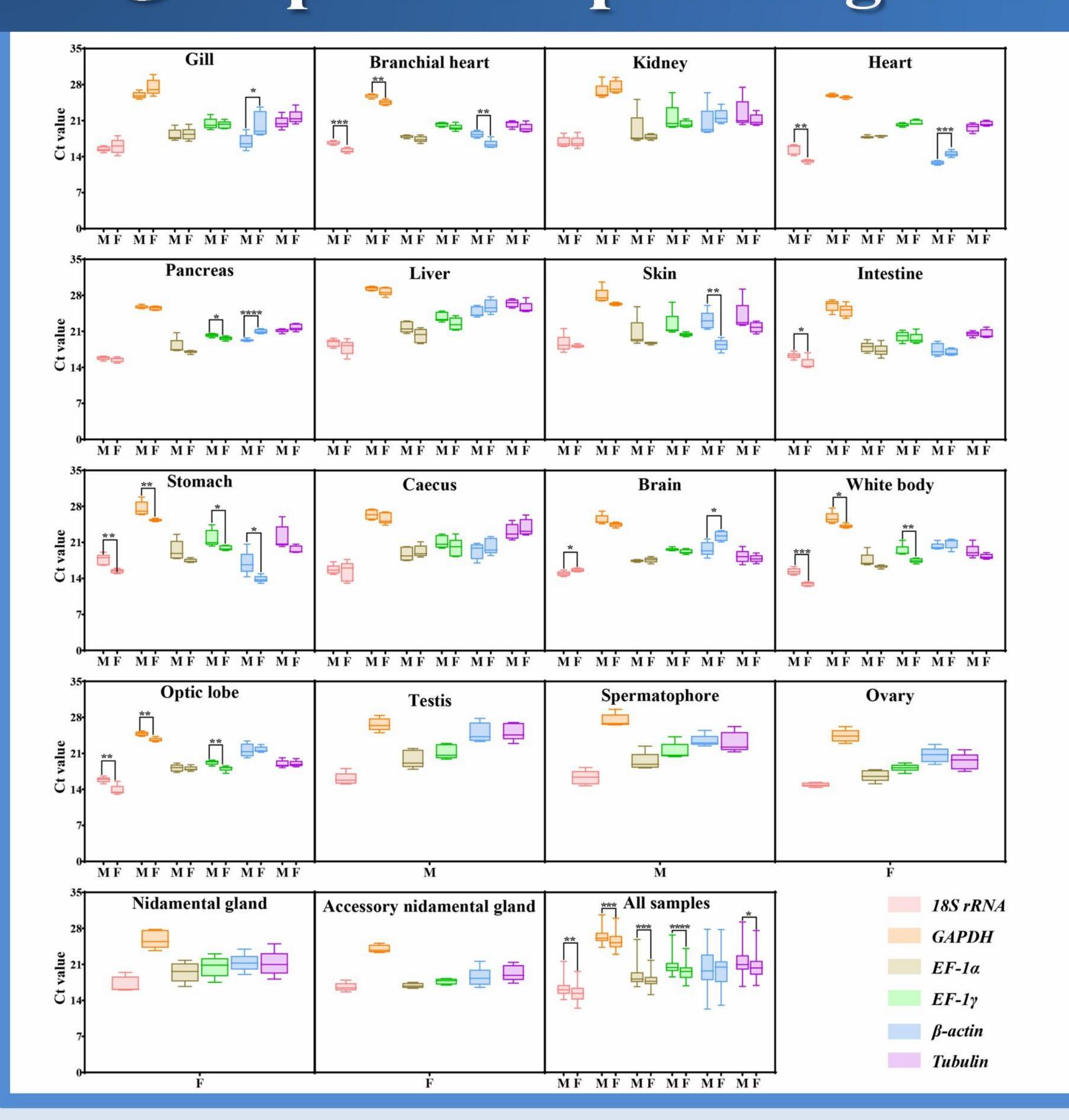


Figure 1. Range of Cycle threshold (Ct) values of the candidate reference genes in different tissues of cuttlefish. Medians are indicated by lines across the box. whisker caps denote the maximum and minimum values. Different colors indicate different candidate genes. M, male. F, female. n = 5. Statistical significance between male and female groups was performed using unpaired two-tailed t-test \*, p < 0.05. \*\*, p < 0.01. \*\*\*, p < 0.001.

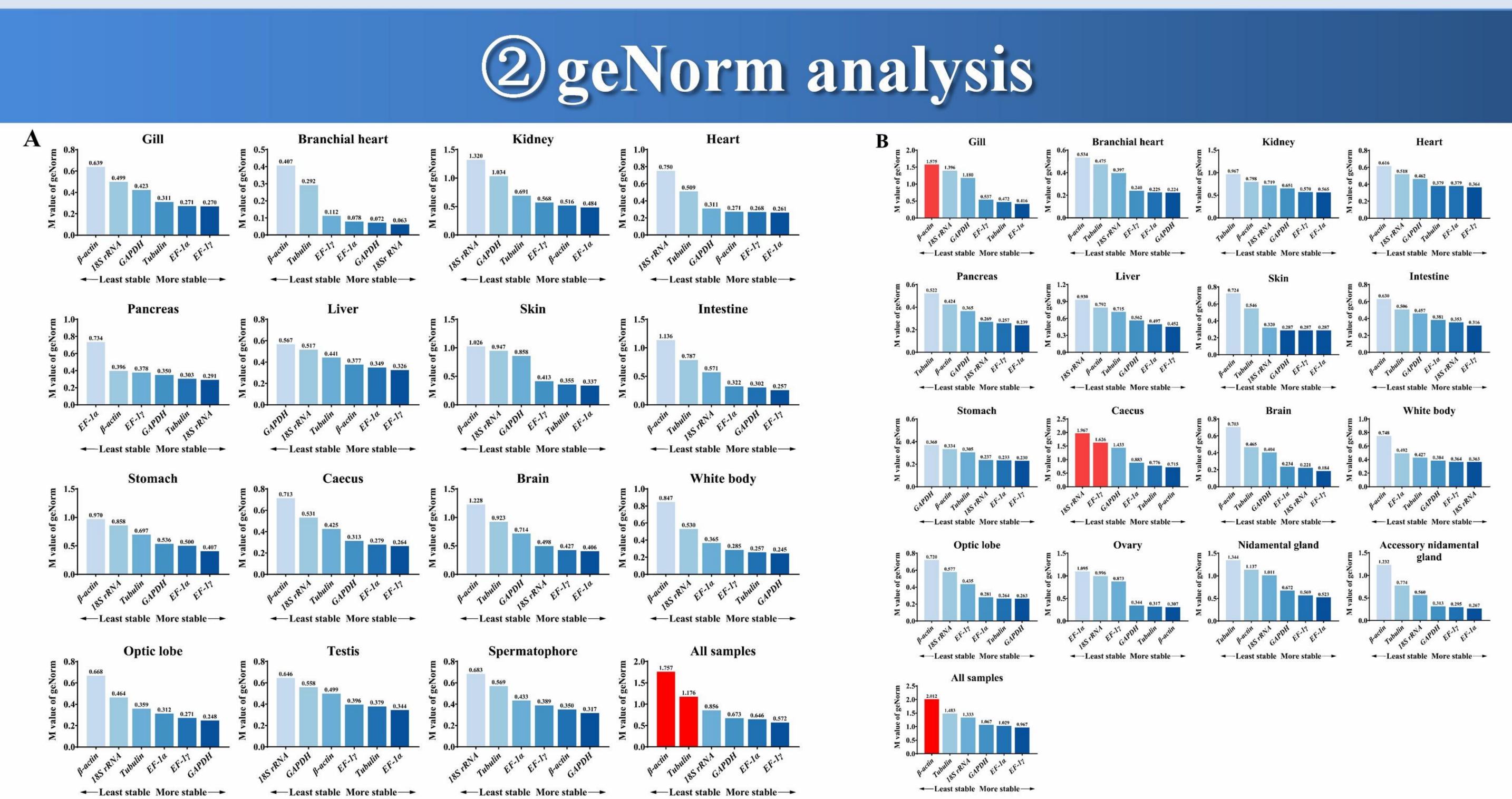


Figure 2. geNorm assessment of reference gene expression stabilities and rankings in in male (A) and female (B) cuttlefish. Six candidate reference genes are ranked according to stability values (M values) assessed by geNorm algorithm. The darker the blue column, the more stable the reference gene. Gene with a M value greater than the cut-off value of 1.5 is not suitable as an internal reference gene, which is highlighted in red column.

#### 3 NormFinder analysis

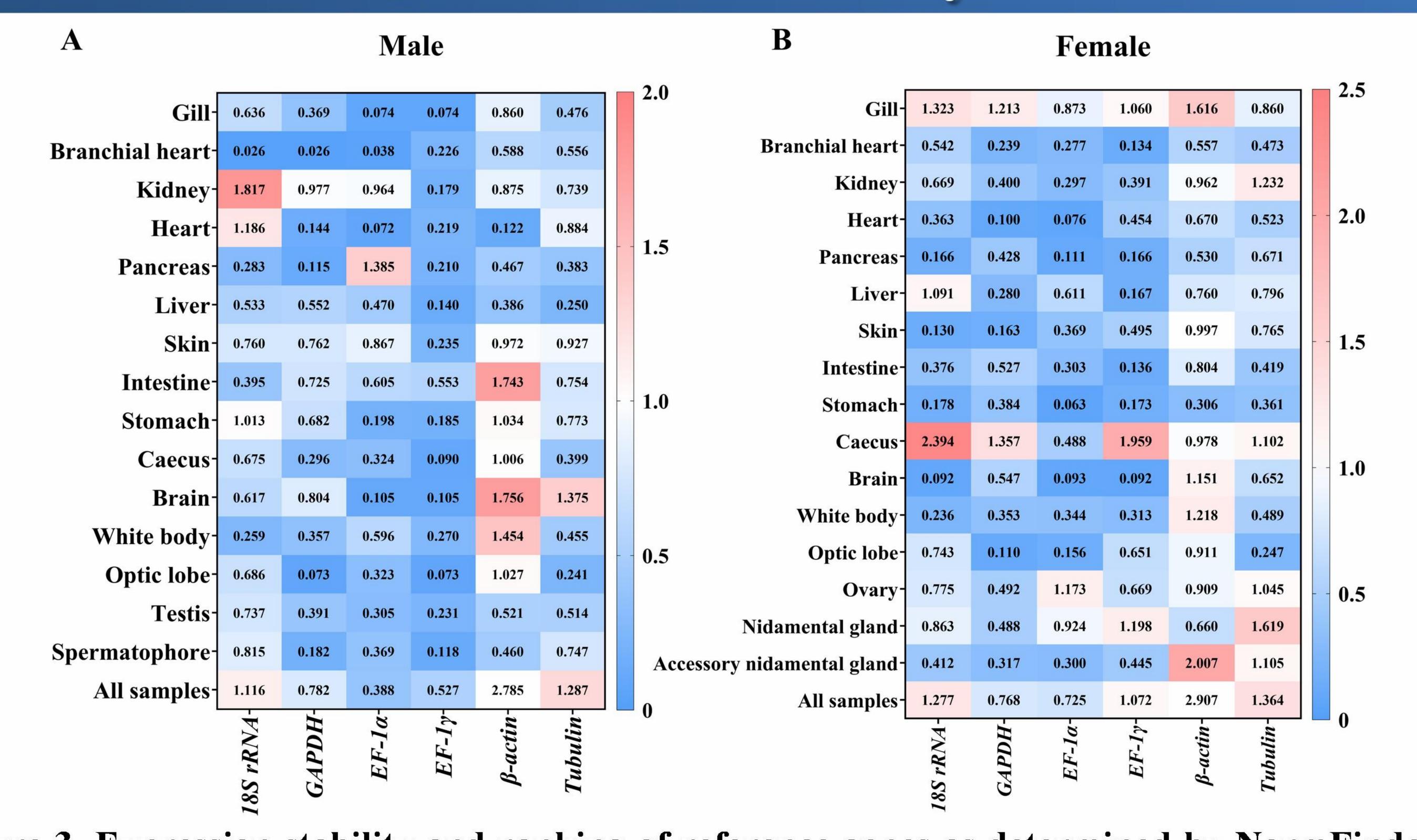


Figure 3. Expression stability and ranking of reference genes as determined by NormFinder in male (A) and female (B) cuttlefish. The stability values of each gene in different tissues are exhibited. Lower values (blue) indicate higher stability, higher values (red) indicate lower stability.

#### 4) BestKeeper analysis

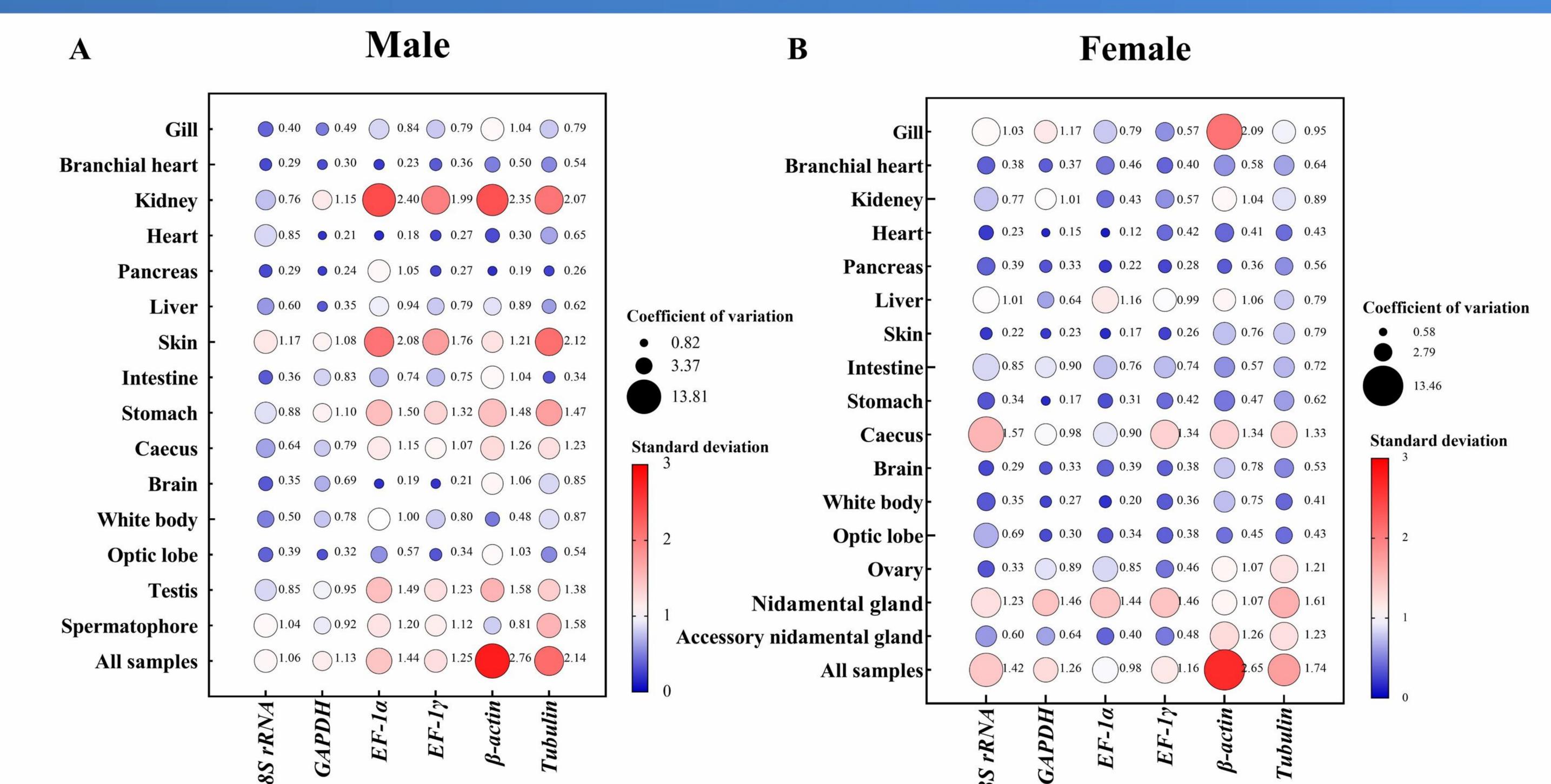


Figure 4. Reference genes stability evaluated by BestKeeper. The standard deviation (SD) values and the standard coefficient of variation (CV) values are marked by colors and the circle size, respectively. The values displayed are SD values. Lower values (blue) and smaller circle indicate higher stability, higher values (red) and larger circle indicate lower stability.

## (5) Comprehensive ranking of reference genes

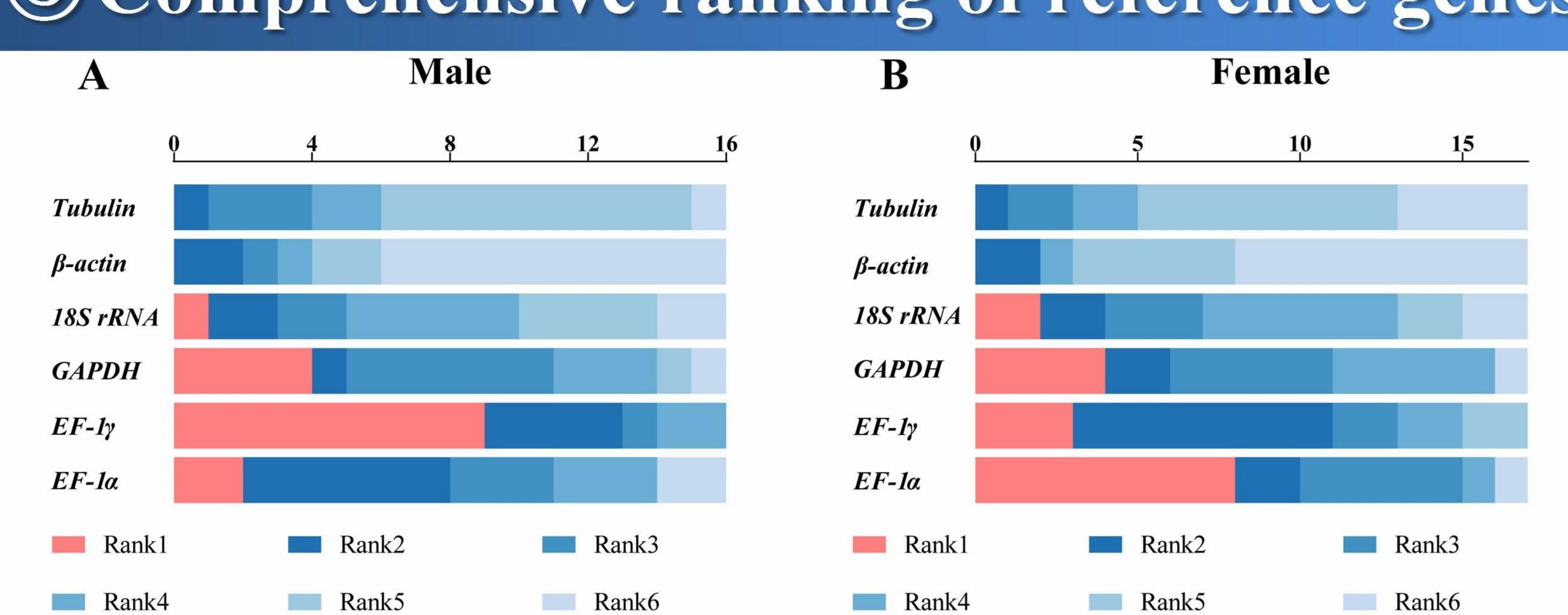


Figure 5. The proportion of each gene across the six rankings in all samples

## **6** The optimal number of internal control genes

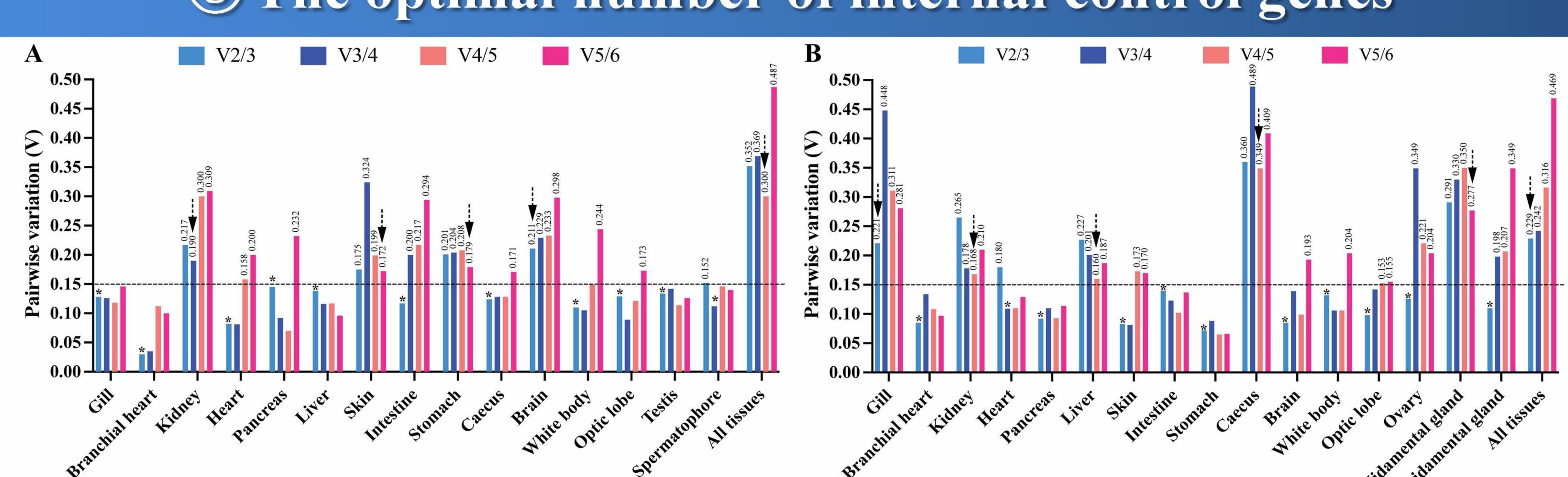


Figure 6. Determination of the optimal number of reference genes for normalization for male (A) and female (B). The dotted line represents the cut-off value of 0.15 for pairwise variation (V) analysis. When Vn/n+1 < 0.15, n indicates the ideal number of reference genes. The optimal number of reference genes for each group is marked with an asterisk. The dashed arrows indicate the Vn/v+1 values are closest to 0.15.

#### Conclusions

Among the six candidate reference genes,  $EF-1\alpha$  and  $EF-1\gamma$  exhibited the most consistent Ct values across different tissues, indicating greater stability compared to  $\beta$ -actin and Tubulin, which showed significant variation. Additionally, except for Tubulin, the Ct values of at least one reference gene differed significantly between male and female cuttlefish within the same tissue. Stability rankings from four analysis methods showed both similarities and differences, but overall,  $EF-1\alpha$  and  $EF-1\gamma$  were the most stable genes in both males and females. In male cuttlefish, the stability ranking was  $EF-1\gamma > EF-1\alpha > GAPDH > 18S \ rRNA > Tubulin > \beta$ -actin, while in females, it was  $EF-1\alpha > EF-1\gamma > GAPDH > 18S \ rRNA > Tubulin > \beta$ -actin. Two reference genes were optimal for most tissues, while three genes were required for the spermatophore and female heart. For tissues with Vn/n+1 values greater than 1.5, the optimal number of reference genes was based on the lowest Vn/n+1 value.